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## REVIEW NEMATODES IN TEA CULTIVATION: PATHOGENESIS AND PLANT DÉFENSE RESPONSES

**Purpose.** To identify the complex relationship between tea plants and important plant-parasitic nematodes, focusing on the severe damage these pests inflict on roots, as well as the sophisticated defense strategies employed by tea plants, particularly their synthesis of protective secondary metabolites to combat nematode attacks

**Results.** Plant-parasitic nematodes pose a serious economic danger to *Camellia sinensis*, the world's most important tea crop, resulting in yield losses of 11–55% and up to \$1 billion yearly. Stunting, wilting, and decreased tea output are caused by around 80 nematode species that harm roots, including *Pratylenchus*, *Radopholus*, *Meloidogyne*, and *Hemicriconemoides*. Due to their immobility, tea plants have developed complex defense mechanisms. These include the formation of nematicidal secondary metabolites (polyphenols, alkaloids, terpenoids), systemic signaling pathways triggered by phytohormones (ethylene, jasmonate, and salicylic acid) that activate defense genes, and physical barriers such as lignin and suberin in cell walls. Nematode control is also aided by beneficial soil bacteria.

**Conclusions.** For sustainable management, it is essential to comprehend relationships. In order to secure the economic future of the tea business, future research should take advantage of natural defences to improve integrated pest control.

**KEYWORDS:** *Camellia sinensis*, plant-parasitic nematodes, plant defense, secondary metabolites, integrated pest management

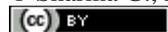
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*Camellia sinensis* (L.) O. Kuntze, commonly known as a tea plant, is a perennial evergreen and a crucial long-term investment for agriculture in tropical and subtropical areas worldwide [1, 2]. Tea first grew in southwestern China

[3] Over the following centuries, people from different cultures introduced it to new parts of the world, and it is now grown in over 60 countries on five continents [4]. Tea is enjoyed by people across all social classes, geographies,

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and cultures. Its widespread appeal stems from its unique flavor and significant health benefits, which are attributed to compounds like tea polyphenols, amino acids, and caffeine [5,6]. Insects and pests are significant factors impacting tea production, leading to yield losses ranging from 11% to 55%. These losses are estimated to result in economic losses amounting to approximately \$500 million to \$1 billion in the tea industry [7]. Among the many factors limiting tea production, plant-parasitic nematodes pose a significant economic threat [8]. The nematode is a significant pathogen, primarily known for devastating tea plantations throughout Asia [9]. The global tea industry faces threats from a diverse range of plant-parasitic nematodes; Sivapalan (1972) [10] recorded over 40 species across 20 genera, while Chen and Chen (1989) [11] reported an even higher number, identifying 82 species linked to tea plants [12].

Plants, being immobile, have evolved defense mechanisms to safeguard themselves against herbivores and microbial infections [13,14]. Due to their immobility, plants face a range of challenges from both living and non-living factors in their environment, including threats from herbivores, pathogens, drought, salinity, UV exposure, temperature extremes, and nutrient deficiencies [15-17]. The perennial and monoculture nature of tea cultivation contributes to substantial annual crop losses. However, geographical variation in pest diversity is evident, influenced by factors such as climate change, altitude, and plantation age [18].

Two types of pests, namely the piercing-sucking pests and chewing pests, are associated with tea [19].

Plants and insects have coexisted for more than 350 million years, leading to the development of unique survival tactics in both groups as they evolved together [14, 20, 21]. Natural selection has enabled plants to survive changing environments by producing diverse secondary metabolites. These metabolites act as a chemical defense mechanism, protecting the plant from pathogen attack and invasion [22]. This adaptive response enables plants to defend against a wide range of threats, from pathogens and herbivores to environmental stresses like drought, salinity, and extreme temperatures [23, 24].

**Classification of Tea pests.** Tea pests can be categorized into three groups based on their location of infestation:

1. Root pests such as nematodes, termites, and cockchafer grubs.
2. Stem pests like the red coffee borer and stem borers.
3. Leaf pests including the tea mosquito bug, thrips, jassids, aphids, flush-worms, looper caterpillars, leaf rollers, and various mite species [25].

This review explores the complex relationship between tea plants and important plant-parasitic nematodes, focusing on the severe damage these pests inflict on roots. It also investigates the sophisticated defense strategies employed by tea plants, particularly their synthesis of protective secondary metabolites to combat nematode attacks

**Root pests:** Pests often affect three layers of plants: the top, midsection, and roots. Pest control often targets the top and intermediate sections of plants, with less emphasis on their roots [26]. Little is known about the interactions of root pests with host plants compared to above-ground insect herbivores [27]. A disturbed soil microflora with limited species diversity and population density of antagonists to fungal root disease agents promotes the spread and incidence of root disease and nematode infections [28]. Deep ploughing and dredging, have been effectively eliminate root pests [29].

**Nematodes:** Nematodes are the most abundant animals on earth and the dominant component of soil [30]. Nematodes are major pests of tea soil in nurseries and new clearings, invading tea seedlings up to 8-9 months old in young plantations. More than 40 species of plant parasitic nematodes from 20 genera have been found in various tea-growing regions across the world [31]. The activity, diversity, and distribution of soil organisms, including nematodes, are strongly influenced by soil properties. Key physical and chemical parameters such as temperature, moisture, water-holding capacity, pH, and electrical conductivity are vital for soil functioning. These factors directly shape nematode community composition and structure [32].

Nematodes have been shown to lower the leaf area and stem girth of tea plants [33]. Infestation spreads mostly through nursery plants on tea farms [25].

Various types of plant parasitic nematodes have been found in tea soils worldwide. Most of these nematodes have not been shown pathogenic (Table. 1). The following species are

known or believed to be pathogenic to tea. Species include *Pratylenchus* spp., *Radopholus similis*, *Meloidogyne* spp., *Hemicriconemoides* sp., *Rotylenchulus reniformis*, *Heliotylenchus* spp., *Paratylenchus curvatus*, *Hoplolaimus* sp. etc.

**Pratylenchus sp:** *Pratylenchus* is a parasite of the tea plant in several tea-producing nations like as, the Philippines, China, Bangladesh, Taiwan, India, Vietnam, the, and Australia [34], Sri Lanka, Japan and United States [35]. This nematode is a major parasite of tea in Iran [35, 36, 37, 38], Japan [39] and Korea [40]. This worm is known as the 'upcountry species of nematode' because of its prevalence in high-elevation regions (2200-3000 m a.s.l.) [41].

**Mode of damage:** Plants respond with apparent symptoms only when a significant portion of their root system is damaged or stops functioning. In extreme circumstances, die-back and death can ensue [25]. The symptoms of tea root-lesion disease caused by *Pratylenchus* spp. infection includes stunting, wilting, yellowing, and a decrease in root length. Such symptoms occur when the nematodes eradicate the root system, resulting in root lesions [8].

**Plant response:** Tea roots have strong cuticle and cell walls, which impede *Pratylenchus* penetration. Infection-trigger Cell walls to reinforced with lignin and suberin, making the plant difficult to penetrate [42]. Tea plants generate secondary metabolites called polyphenols, which have nematicidal characteristics and impede *Pratylenchus* development and reproduction [43]. Tea plants develop a systemic response to *Pratylenchus* infection by activating defense genes and producing signalling molecules such as salicylic acid [44]. Phytohormones like ethylene and jasmonate also resist the *Pratylenchus* infection [45]. Tea plants interact with helpful bacteria in the soil, helping to reduce *Pratylenchus* numbers [46].

**Radopholus similis:** The species was initially identified as a tea pest in Java, Indonesia [47]. *R. similis* was detected in three locations: Bangka, Gambung in West Java, and on tea plantations elsewhere in Java [48]. Steiner and Buhner (1933) found tea to be a suitable host for this nematode. it is found in Africa, Asia, North, and South America, Australia, and a few European countries [49]. The *Radopholus similis* was reported from 50m to 1000m altitude [50].

**Mode of damage:** Tea plants infested with *R. similis* exhibit symptoms similar to those produced by *Pratylenchus loosi*, including

stunting with twiggy branches, defoliation, early blooming, and fruiting. Infested plants have scant and dry roots, as opposed to healthy plants' white, succulent feeder roots. *R. similis* causes lesions on tea roots, but they are much less than those caused by *P. loosi* [51].

**Plant response:** Plants generate secondary metabolites, which help them defend against worm infections. Secondary metabolites mostly consist of alkaloids, terpenoids, and phenylpropanoids [52]. Due to nematode infection, plants accumulate toxins, modify cell wall, and synthesize phytoalexins [53].

**Meloidogyne sp:** Root-knot nematodes (RKNs), *Meloidogyne* spp., are among the oldest and most economically important plant-parasitic nematodes (PPNs). They are serious pests in agricultural production, causing annual global losses of USD 157 billion. [54, 55] *M. javanica*, *M. incognita*, and *M. brevicaua* are common root-knot nematodes (RKN) found in tea plants, particularly in India [33] and tea growing regions of Africa [56].

**Mode of damage:** *Meloidogyne* enters the root and reaches the growing vascular strands at the elongation zone, resulting in many mitotic cycles without cytokinesis, which leads them to expand and become multinucleated (up to >100 nuclei), leading to the creation of "galls" or "root knots" [57]. Several physiological and morphological changes occur during the formation of root knots and turn them into nematode-feeding sites, which give nutrients for the worm's growth [58]. Seedling plants are more vulnerable to root injury (Tabl.) than vegetatively produced clonal tea plants of the same age, perhaps due to their smaller root mass [59].

**Plant response:** When RKNs infect a host plant, the plant initiates a defense response, both naturally and artificially, through the coordinated expression of multiple plant signaling pathways, including phytohormones such as salicylic acid, ethylene, and jasmonic acid, pathogenesis-related (PR) proteins, and various plant transcription factors. The outcomes of these highly coordinated signaling responses eventually determine the susceptibility or resistance of plants to RKNs [73]. Resistance to the root-knot nematode can be attributed to the hypersensitive response (HR) and higher phenolic levels, which result in barrier deposition (e.g., lignin) [74].

**Hemicriconemoides sp:** These nematodes, commonly known as 'sheathoid' nemato-

Table

Tea root Nematodes: Host Spectrum, Species Diversity, and host Interactions

Sn	Name	Distribution	No of species	Host range	Symptom	Host defence
1	<i>Pratylenchus sp.</i>	Worldwide except. Antarctica [60]	100 species [61]	400 plant species (ornamental, vegetable, tree etc) [62]	Root development disruption, poorly developed foliage [60]	Polyphenols, salicylic acid, Phytohormones like ethylene and jasmonate
2	<i>Radopholus sp.</i>	Worldwide mostly tropical and subtropical areas [63]	Around 20 Species [64]	250 plant species [65] like banana, citrus, pepper, coffee, Cucumber [63], tea [66]	Stunting with twiggy branches, defoliation, early blooming, and fruiting [51]	Alkaloids, terpenoids, phenylpropanoids and phytoalexins
3	<i>Meloidogyne sp.</i>	Worldwide [67]	Around 100 species [68]	Broad host range [69]	Gall or root-knot, effect root system, and significant yield losses [67]	Salicylic acid, ethylene, jasmonic acid, Phenolic Compound
4	<i>Hemicriconemoides sp.</i>	Worldwide [70]	54 Species [71,72]	Higher plants, primarily woody perennials, and vines [70]	Root rot, leaf necrosis, suppressed root growth, lesions, and coarse roots [70]	Alkaloids, terpenoids, and phenylpropanoids

des due to their smooth-annulated outer accessory sheath, are reported worldwide. They are associated with various higher plants, primarily woody perennials, and vines [68]. *Hemicriconemoides* spp. (Sheathoid nematodes) are sexually dimorphic plant-parasitic nematodes found mostly in warm areas [75]. The genus *Hemicriconemoides* consists of 54 species [71, 72]. *Hemicriconemoides* is a significant nematode pest of tea in Japan [39]. The nematode has been detected in Taiwan [10] and Iran [76].

**Mode of damage:** Common symptoms of infestation by these nematodes include root rot, leaf necrosis, suppressed root growth, le-

sions, and coarse roots [65, 62]. This ectoparasitic nematode feeds only on tea feeder roots. Continuous feeding by this worm causes the sloughing off of the root cortex, producing a brownish discoloured stele [77]. Plants afflicted with these parasites may exhibit symptoms such as stunting, early wilting, leaf yellowing, root deformity, necrosis of cortical root tissues, and nutritional shortages [78].

**Plant response:** Plants produce secondary metabolites that help them defend against helminth infections. Secondary metabolites mainly consist of alkaloids, terpenoids, and phenylpropanoids [43].

### Conclusion

Globally, tea production and economic value (*Camellia sinensis*) are damaged and tea production reduced up to 11-55%, with some estimates at \$1 billion loss, due to yield loss and production reductions as a result of plant-parasitic nematodes. Pathogenic nematodes, including *Pratylenchus*, *Radopholus*, *Meloidogyne*,

and *Hemicriconemoides*, have a lasting and damaging effect, along with wilting and reduced tea production.

Due to these challenges, the tea plant, and by extension the immobilized plant, has come up with a few layers of the defense strategy. The first part of the defense strategy is the

production of polyphenols, alkaloids, and terpenoids, which are nematicidal, and the activation of 'systemic' defense along the pathways of the 'phytohormones' ethylene, jasmonate, and salicylic acid, and the lignin and suberin pathways. Defense interactions at this level are further supported by the physical structures and beneficial microbiomes in the soil.

To develop effective and sustainable management approaches, plant-nematode interactions must be understood. Future tea production will be building this 'plant intelligence' and boosting the 'defenses.' Added these to the rest of the Integrated Pest Management (IPM) tea systems, the chance of reduced chemical dependence and economic viability increases and remains sustainable in production worldwide.

### Conflict of Interest

The authors declare no conflict of interest regarding the publication of this manuscript. Furthermore, the authors have fully adhered to ethical norms, including avoiding plagiarism, data falsification, and duplicate publication.

**Authors Contribution:** all authors have contributed equally to this work.

In this study, generative artificial intelligence was used only to help find and compile recent academic literature on the subject. The authors, who take full responsibility for the work's intellectual content, personally performed the crucial tasks of data analysis, information synthesis, and drawing scientific conclusions.

### References

1. Zou, Y., Shen, F., Zhong, Y., Lv, C., Pokharel, S. S., Fang, W., & Chen, F. (2022). Impacts of intercropped maize ecological shading on tea foliar and functional components, insect pest diversity and soil microbes. *Plants*, 11(14), 1883. <https://doi.org/10.3390/plants11141883>
2. Han, Z., Zhang, C., Zhang, H., Duan, Y., Zou, Z., Zhou, L., ... & Ma, Y. (2022). CsMYB transcription factors participate in jasmonic acid signal transduction in response to cold stress in tea plant (*Camellia sinensis*). *Plants*, 11(21), 2869. <https://doi.org/10.3390/plants11212869>
3. Xia, E., Tong, W., Hou, Y., An, Y., Chen, L., Wu, Q., ... & Wan, X. (2020). The reference genome of tea plant and resequencing of 81 diverse accessions provide insights into its genome evolution and adaptation. *Molecular plant*, 13(7), 1013-1026. <https://doi.org/10.1016/j.molp.2020.04.010>
4. Yue, C., Peng, H., Li, W., Tong, Z., Wang, Z., & Yang, P. (2022). Untargeted metabolomics and transcriptomics reveal the mechanism of metabolite differences in spring tender shoots of tea plants of different ages. *Foods*, 11(15), 2303. <https://doi.org/10.3390/foods11152303>
5. Chen, Y., Cheng, S., Dai, J., Wang, L., Xu, Y., Peng, X., ... & Peng, C. (2021). Molecular mechanisms and applications of tea polyphenols: A narrative review. *Journal of Food Biochemistry*, 45(10), e13910. <https://doi.org/10.1111/jfbc.13910>
6. Wang, W., Zhou, X., Hu, Q., Wang, Q., Zhou, Y., Yu, J., ... & Li, X. (2025). Lignin Metabolism Is Crucial in the Plant Responses to *Tambocerus elongatus* (Shen) in *Camellia sinensis* L. *Plants*, 14(2), 260. <https://doi.org/10.3390/plants14020260>
7. Kachhawa, D., & Kumawat, K. (2018). *Oligonychus coffeae*: red spider mite of tea: a review. *J Entomol Zool Stud*, 6(3), 519-524. <https://www.academia.edu/download/89555883/6-3-2-927.pdf>
8. Rahanandeh, H., Khodakaramian, G., Hassanzadeh, N., Seraji, A., Asghari, S. M., & Tarang, A. R. (2012). Inhibition of tea root lesion nematode, *Pratylenchus loosi*, by rhizosphere bacteria. [https://www.researchgate.net/publication/339540753\\_Inhibition\\_of\\_Tea\\_Root\\_Lesion\\_Nematode\\_Pratylenchus\\_Loosi\\_by\\_rhizosphere\\_bacteria](https://www.researchgate.net/publication/339540753_Inhibition_of_Tea_Root_Lesion_Nematode_Pratylenchus_Loosi_by_rhizosphere_bacteria)
9. EFSA Panel on Plant Health (PLH), Bragard, C., Baptista, P., Chatzivassiliou, E., Di Serio, F., Gonthier, P., ... & Reignault, P. L. (2024). Pest categorisation of *Pratylenchus loosi*. *EFSA Journal*, 22(1), e8548. <https://doi.org/10.2903/j.efsa.2024.8548>
10. Sivapalan, P. (1972). Nematode pests of tea. *Nematode pests of tea*. <https://www.cabidigitallibrary.org/doi/full/10.5555/19722001016>
11. Chen, Z. M., & Chen, X. F. (1989). An analysis of world tea pest fauna. *J. Tea Sci*, 9(1), 13-22. <http://www.tscha.org/CN/abstract/abstract16.shtml>
12. Paul, S. K., Ahmed, M., & Mamun, M. S. A. (2014). Biopesticides: A potential tool for the management of plant parasitic nematodes in tea. *Tea Journal of Bangladesh*, 43, 24-33. [https://www.bjstea.org/tjb\\_vol\\_43\\_2014/5.%2520Biopesticides-%2520A%2520Potential%2520Tool%2520for%2520the%2520Management%2520of%2520Plant%2520Parasitic%2520Nematodes%2520in%2520Tea.pdf](https://www.bjstea.org/tjb_vol_43_2014/5.%2520Biopesticides-%2520A%2520Potential%2520Tool%2520for%2520the%2520Management%2520of%2520Plant%2520Parasitic%2520Nematodes%2520in%2520Tea.pdf)
13. Wink, M. (2008). Plant secondary metabolism: diversity, function and its evolution. *Natural Product Communications*, 3(8), 1934578X0800300801. <https://doi.org/10.1177/1934578X0800300801>

14. Petschenka, G., Halitschke, R., Züst, T., Roth, A., Stiehler, S., Tenbusch, L., ... & Exnerová, A. (2022). Sequestration of defences against predators drives specialized host plant associations in preadapted milkweed bugs (Heteroptera: Lygaeinae). *The American Naturalist*, 199(6), E211-E228. <https://doi.org/10.1086/719196>
15. Freeman, B., & Beattie, G. (2008). An overview of plant defences against pathogens and herbivores. <https://doi.org/10.1094/PHI-I-2008-0226-01>
16. Jan, R., Asaf, S., Numan, M., & Kim, K. M. (2021). Plant secondary metabolite biosynthesis and transcriptional regulation in response to biotic and abiotic stress conditions. *Agronomy*, 11 (5): 968. <https://doi.org/10.3390/agronomy11050968>
17. Teklić, T., Parađiković, N., Špoljarević, M., Zeljković, S., Lončarić, Z., & Lisjak, M. (2021). Linking abiotic stress, plant metabolites, biostimulants and functional food. *Annals of Applied Biology*, 178(2), 169-191. <https://doi.org/10.1111/aab.12651>
18. Deka, B., Babu, A., Baruah, C., & Barthakur, M. (2021). Nanopesticides: A systematic review of their prospects with special reference to tea pest management. *Frontiers in Nutrition*, 8, 686131. <https://doi.org/10.3389/fnut.2021.686131>
19. Naskar, S., Roy, C., Ghosh, S., Mukhopadhyay, A., Hazarika, L. K., Chaudhuri, R. K., ... & Chakraborti, D. (2021). Elicitation of biomolecules as host defense arsenals during insect attacks on tea plants (*Camellia sinensis* (L.) Kuntze). *Applied microbiology and biotechnology*, 1-13. <https://doi.org/10.1007/s00253-021-11560-z>
20. War, A. R., Paulraj, M. G., Ahmad, T., Buhroo, A. A., Hussain, B., Ignacimuthu, S., & Sharma, H. C. (2012). Mechanisms of plant defense against insect herbivores. *Plant signaling & behavior*, 7(10), 1306-1320. <https://doi.org/10.4161/psb.21663>
21. Sharma, G., Kumar, A., Naushad, M., Al-Misned, F. A., El-Serehy, H. A., Ghfar, A. A., ... & Stadler, F. J. (2020). Graft copolymerization of acrylonitrile and ethyl acrylate onto *Pinus roxburghii* wood surface enhanced physicochemical properties and antibacterial activity. *Journal of Chemistry*, May, 1-16. <https://doi.org/10.1155/2020/6285354>
22. Neves, D., Figueiredo, A., Maia, M., Laczko, E., Pais, M. S., & Cravador, A. (2023). A Metabolome Analysis and the Immunity of *Phlomis purpurea* against *Phytophthora cinnamomi*. *Plants*, 12(10), 1929. <https://doi.org/10.3390/plants12101929>
23. Koza, N. A., Adedayo, A. A., Babalola, O. O., & Kappo, A. P. (2022). Microorganisms in plant growth and development: roles in abiotic stress tolerance and secondary metabolites secretion. *Microorganisms*, 10(8), 1528. <https://doi.org/10.3390/microorganisms10081528>
24. Simsek, M., & Whitney, K. (2024). Examination of primary and secondary metabolites associated with a plant-based diet and their impact on human health. *Foods*, 13(7), 1020. <https://doi.org/10.3390/foods13071020>
25. Mamun, M. S. A., Ahmed, M., & Paul, S. K. (2011). Control of plant parasitic nematodes of tea soil using different species of green crops in Bangladesh. *Tea Journal of Bangladesh*, 40, 1-7. [https://www.bjstea.org/tjb\\_vol\\_40\\_2011/1.%2520Control%2520of%2520Plant%2520Parasitic%2520Nematodes%2520of%2520Tea%2520Soil%2520Using%2520Different%2520Species%2520of%2520Green%2520Crops%2520in%2520Bangladesh.pdf](https://www.bjstea.org/tjb_vol_40_2011/1.%2520Control%2520of%2520Plant%2520Parasitic%2520Nematodes%2520of%2520Tea%2520Soil%2520Using%2520Different%2520Species%2520of%2520Green%2520Crops%2520in%2520Bangladesh.pdf)
26. Huang, L., Zhuang, Y., & Liu, Q. (2023). A mathematical model study on plant root pest management. *AIMS Mathematics*, 8(4), 9965-9981. <https://doi.org/10.3934/math.2023504>
27. Johnson, S. N., Crawford, J. W., Gregory, P. J., Grinev, D. V., Mankin, R. W., Masters, G. J., ... & Zhang, X. (2007). Non-invasive techniques for investigating and modelling root-feeding insects in managed and natural systems. *Agricultural and Forest Entomology*, 9(1), 39-46. <https://doi.org/10.1111/j.1461-9563.2006.00315.x>
28. Zhou, D., Feng, H., Schuelke, T., De Santiago, A., Zhang, Q., Zhang, J., ... & Wei, L. (2019). Rhizosphere microbiomes from root knot nematode non-infested plants suppress nematode infection. *Microbial Ecology*, 78(2), 470-481. <https://doi.org/10.1007/s00248-019-01319-5>
29. Márquez, M. (2001). Efecto de la mecanización sobre la población de plagas de la raíz en caña de azúcar y su estimación con diferentes tamaños de unidad de muestreo. In: *Memoria X Congreso Nacional de la Caña de Azúcar y II Simposio Nacional de Plagas*. Guatemala, ATAGUA. (pp. 15-20). <https://dialnet.unirioja.es/descarga/libro/572719.pdf#page=217>
30. Van Den Hoogen, J., Geisen, S., Routh, D., Ferris, H., Traunspurger, W., Wardle, D. A., ... & Crowther, T. W. (2019). Soil nematode abundance and functional group composition at a global scale. *Nature*, 572(7768), 194-198. <https://doi.org/10.1038/s41586-019-1418-6>
31. Monjil, M. S. (2024). Current status of nematode problems of crops in Bangladesh. *Nematode Problems in Crops and their Management in South Asia*, 73. [https://www.researchgate.net/profile/Shanowly-Mondal-Ghosh/publication/384253890\\_Nematode\\_Problems\\_in\\_Crops\\_and\\_their\\_Management\\_in\\_South\\_Asia\\_hard\\_man\\_v1\\_1/links/66f14c6119c9496b1fb7ea77/Nematode-Problems-in-Crops-and-their-Management-in-South-Asia-hard-man-v1-1.pdf](https://www.researchgate.net/profile/Shanowly-Mondal-Ghosh/publication/384253890_Nematode_Problems_in_Crops_and_their_Management_in_South_Asia_hard_man_v1_1/links/66f14c6119c9496b1fb7ea77/Nematode-Problems-in-Crops-and-their-Management-in-South-Asia-hard-man-v1-1.pdf)
32. Al-Ghamdi, A. A. M. (2021). Relationship between nematodes and some soil properties in the rhizosphere of banana plants. *International Letters of Natural Sciences*, 82. <https://doi.org/10.18052/www.scipress.com/ILNS.82.1>

33. Orisajo, S. B. (2012). Distribution of plant-parasitic nematodes associated with tea in Nigeria. *World journal of Agricultural sciences*, 8(5), 459-463. [https://www.idosi.org/ajbas/ajbas5\(4\)13/5.pdf](https://www.idosi.org/ajbas/ajbas5(4)13/5.pdf)
34. EFSA Panel on Plant Health (PLH), Bragard, C., Baptista, P., Chatzivassiliou, E., Di Serio, F., Gonthier, P., ... & Reignault, P. L. (2024). Pest categorisation of *Pratylenchus loosi*. *EFSA Journal*, 22(1), e8548. <https://doi.org/10.2903/j.efsa.2024.8548>
35. Amarasena, P. G. D. S., Mohotti, K. M., De Costa, D. M., & Fosu-Nyarko, J. (2020). Morphometric and molecular characterization of isolates of the root lesion nematode, *Pratylenchus loosi* infecting tea in Sri Lanka. *Tropical Agricultural Research*, 31(1). <https://doi.org/10.4038/tar.v31i1.8344>
36. Pourjam, E., Kheiri, A., & Geraert, E. (1997). The genus *Pratylenchus* Filip'jev, 1936 (Tylenchida: Pratylenchidae) from North of Iran. The genus *Pratylenchus* Filip'jev, 1936 (Tylenchida: Pratylenchidae) from North of Iran., 62(3a), 741-756. <https://www.cabidigitallibrary.org/doi/full/10.5555/19981700196>
37. Pourjame, E., Waeyenberge, L., Moens, M., & Geraert, E. (1999). Morphological, morphometrical and molecular study of *Pratylenchus coffeae* and *P. loosi* (Nematoda: Pratylenchidae). <https://www.cabidigitallibrary.org/doi/full/10.5555/20001701238>
38. Seraji, A., Pourjam, E., Tanha, M. Z., & Safaei, N. (2007). Biology and population dynamics of tea root lesion nematode (*Pratylenchus loosi*) in Iran. <https://sid.ir/paper/48216/en>
39. Kaneko, T., & Ichinohe, M. (1968). Notes on the nematode species and their bionomics associated with tea roots in Japan. *Review of Plant Protection Research*, 1, 122-124. <https://www.cabidigitallibrary.org/doi/full/10.5555/19670801585>
40. Park ByeongYong, P. B., Choi DongRo, C. D., Lee JaeKook, L. J., Choi YoungEoun, C. Y., & Shin GilHo, S. G. (2002). An unrecorded species of *Pratylenchus loosi* Loof (Tylenchida: Pratylenchidae) from tea in Korea. <https://koreascience.kr/article/JAKO200211921497549.pdf>
41. Kidane, S. A., Meressa, B. H., Haukeland, S., Hvoslef-Eide, T., Magnusson, C., Couvreur, M., ... & Coyne, D. L. (2020). Occurrence of plant-parasitic nematodes on enset (*Ensete ventricosum*) in Ethiopia with focus on *Pratylenchus goodeyi* as a key species of the crop. *Nematology*, 23(5), 529-541 <https://doi.org/10.1163/15685411-bja10058>
42. Vilas, J. M., Romero, F. M., Rossi, F. R., Marina, M., Maiale, S. J., Calzadilla, P. I., ... & Gárriz, A. (2018). Modulation of plant and bacterial polyamine metabolism during the compatible interaction between tomato and *Pseudomonas syringae*. *Journal of plant physiology*, 231, 281-290. <https://doi.org/10.1016/j.jplph.2018.09.014>
43. Barbosa, P., Faria, J. M., Cavaco, T., Figueiredo, A. C., Mota, M., & Vicente, C. S. (2024). Nematicidal activity of phytochemicals against the root-lesion nematode *Pratylenchus penetrans*. *Plants*, 13(5), 726. <https://doi.org/10.3390/plants13050726>
44. Abd-Elgawad, M. M. (2021). Optimizing safe approaches to manage plant-parasitic nematodes. *Plants*, 10(9), 1911. <https://doi.org/10.3390/plants10091911>
45. Sikder, M. M., & Vestergård, M. (2020). Impacts of root metabolites on soil nematodes. *Frontiers in plant science*, 10, 1792. <https://doi.org/10.3389/fpls.2019.01792>
46. Rahanandeh, H., Khodakaramian, G., Hassanzadeh, N., Seraji, A., & Asghari, S. M. (2013). Evaluation of antagonistic *Pseudomonas* against root lesion nematode of tea. <http://dx.doi.org/10.12692/ijb/3.3.32-40>
47. Campos, V. P., Sivapalan, P., & Gnanapragasam, N. C. (1990). Nematode parasites of coffee, cocoa and tea. <https://www.cabidigitallibrary.org/doi/full/10.5555/19901146381>
48. Djiwanti, S. R. (2019, March). Taxa status of some reported plant parasitic nematodes in Indonesia. In *IOP Conference Series: Earth and Environmental Science*, 250(1), 012100. IOP Publishing. <https://doi.org/10.1088/1755-1315/250/1/012100>
49. Kuhlmann, U., Jenner, W., & Reeder, R. (2023). PlantwisePlus Annual Report 2022. [https://www.plantwise.org/Uploads/Plantwise/PlantwisePlus/PlantwisePlus\\_Annual\\_Report\\_2022.pdf](https://www.plantwise.org/Uploads/Plantwise/PlantwisePlus/PlantwisePlus_Annual_Report_2022.pdf)
50. Hulupi, R., Mulyadi, M., & Andayani, B. (2006). Distribution of *Radopholus similis* and *Pratylenchus Coffeae* Nematodes in Coffee Plantation. *Pelita Perkebunan*, 23(3), 158458. <https://doi.org/10.22302/iccricri.jur.pelita-perkebunan.v23i3.41>
51. Gnanapragasam, N. C. (1983). Incidence of nematode damage in the mid-country caused by the burrowing nematode, *Radopholus similis*. *Tea Quarterly*, 52(1). <https://sljournals.tripod.com/tea-quarterly-abstracts/tq-abstracts-v52-n1.html#gnanapragasam>
52. Chitwood, D. J. (2003). Nematicides. *Encyclopedia of agrochemicals*. <https://doi.org/10.1002/047126363X.agr171>
53. Appel, H. M. (1993). Phenolics in ecological interactions: the importance of oxidation. *Journal of chemical ecology*, 19, 1521-1552. <https://doi.org/10.1038/nbt.1482>
54. Abad, P., Gouzy, J., Aury, J. M., Castagnone-Sereno, P., Danchin, E. G., Deleury, E., ... & Wincker, P. (2008). Genome sequence of the metazoan plant-parasitic nematode *Meloidogyne incognita*. *Nature biotechnology*, 26(8), 909-915. <https://doi.org/10.1038/nbt.1482>

55. Rusinque, L., Camacho, M. J., Serra, C., Nobrega, F., & Inacio, M. L. (2023). Root-knot nematode assessment: species identification, distribution, and new host records in Portugal. *Frontiers in Plant Science*, 14, 1230968. <https://doi.org/10.3389/fpls.2023.1230968>
56. Kamunya, S. M., Lang'at, J. K., Muoki, R. C., Nyabundi, K., Wachira, F. N., Otieno, W., & Sudoi, V. (2008). Performance of tea clones established in root knot nematode host spots under varying treatment regimes. <https://www.cabidigitallibrary.org/doi/full/10.5555/20103355298>
57. Smant, G., Helder, J., & Govere, A. (2018). Parallel adaptations and common host cell responses enabling feeding of obligate and facultative plant parasitic nematodes. *The Plant Journal*, 93(4), 686-702. <https://doi.org/10.1111/tpj.13811>
58. Palomares-Rius, J. E., Escobar, C., Cabrera, J., Vovlas, A., & Castillo, P. (2017). Anatomical alterations in plant tissues induced by plant-parasitic nematodes. *Frontiers in plant science*, 8, 288614. <https://doi.org/10.3389/fpls.2017.01987>
59. Gnanapragasam, N. C., & Mohotti, K. M. (2018). Nematode parasites of tea. In *Plant parasitic nematodes in subtropical and tropical agriculture* (pp. 584-616). Wallingford UK: CAB International. <https://doi.org/10.1079/9781786391247.0584>
60. Kantor, C., Eisenback, J. D., & Kantor, M. (2024). Biosecurity risks to human food supply associated with plant-parasitic nematodes. *Frontiers in Plant Science*, 15, 1404335. <https://doi.org/10.3389/fpls.2024.1404335>
61. Handoo, Z. A., Skantar, A. M., Kantor, M. R., Hafez, S. L., & Hult, M. N. (2020). Molecular and morphological characterization of the amaryllis lesion nematode, *Pratylenchus hippeastri* (Inserra et al., 2007), from California. *Journal of Nematology*, 52, e2020-58. <https://doi.org/10.21307/jofnem-2020-058>
62. Figueiredo, J., Vieira, P., Abrantes, I., & Esteves, I. (2022). Commercial potato cultivars exhibit distinct susceptibility to the root lesion nematode *pratylenchus penetrans*. *Horticulturae*, 8(3), 244. <https://doi.org/10.3390/horticulturae8030244>
63. Bahadur, A. (2022). Vegetables Crops in India. Nematodes: Recent advances, management and new perspectives, 3. <https://www.intechopen.com/chapters/77474>
64. Trinh, P. Q., Nguyen, C. N., Waeyenberge, L., Subbotin, S. A., Karssen, G., & Moens, M. (2004). *Radopholus arabocoffeae* sp. n. (Nematoda: Pratylenchidae), a nematode pathogenic to *Coffea arabica* in Vietnam, and additional data on *R. duriophilus*. *Nematology*, 6(5), 681-693. <https://doi.org/10.1163/1568541042843577>
65. Bernard, G. C., Egnin, M., & Bonsi, C. (2017). The impact of plant-parasitic nematodes on agriculture and methods of control. In *Nematology-concepts, diagnosis and control*. IntechOpen. <https://doi.org/10.1088/1755-1315/250/1/012100>
66. Djiwanti, S. R. (2019, March). Taxa status of some reported plant parasitic nematodes in Indonesia. In *IOP Conference Series: Earth and Environmental Science* (Vol. 250, No. 1, p. 012100). IOP Publishing <https://doi.org/10.1088/1755-1315/250/1/012100>
67. Rusinque, L., Camacho, M. J., Serra, C., Nobrega, F., & Inacio, M. L. (2023). Root-knot nematode assessment: species identification, distribution, and new host records in Portugal. *Frontiers in Plant Science*, 14, 1230968. <https://doi.org/10.3389/fpls.2023.1230968>
68. Subbotin, S. A., Rius, J. E. P., & Castillo, P. (2021). Systematics of root-knot nematodes (Nematoda: Meloidogynidae) (Vol. 14). Brill. [https://books.google.co.in/books?hl=en&lr=&id=xs87EAAAQBAJ&oi=fnd&pg=PR3&dq=76.%09Subbotin,+S.+A.,+Rius,+J.+E.+P.,+%26+Castillo,+P.+\(2021\).+Systematics+of+root-knot+nematodes+\(Nematoda:+Meloidogynidae\)+\(Vol.+14\).+Brill&ots=g2eW0hg5SX&sig=CiExGlpd4A8q541lB9oXktn8gy8&redir\\_esc=y#v=onepage&q&f=false](https://books.google.co.in/books?hl=en&lr=&id=xs87EAAAQBAJ&oi=fnd&pg=PR3&dq=76.%09Subbotin,+S.+A.,+Rius,+J.+E.+P.,+%26+Castillo,+P.+(2021).+Systematics+of+root-knot+nematodes+(Nematoda:+Meloidogynidae)+(Vol.+14).+Brill&ots=g2eW0hg5SX&sig=CiExGlpd4A8q541lB9oXktn8gy8&redir_esc=y#v=onepage&q&f=false)
69. Jones, J. T., Haegeman, A., Danchin, E. G., Gaur, H. S., Helder, J., Jones, M. G., ... & Perry, R. N. (2013). Top 10 plant-parasitic nematodes in molecular plant pathology. *Molecular plant pathology*, 14(9), 946-961. <https://doi.org/10.1111/mpp.12057>
70. Sharma, H., & Chaubey, A. K. (2023). Molecular and phenotypic characterization of *Hemicriconemoides rosae* (Rathour et al., 2003) from mustard rhizosphere in India. *The Journal of Basic and Applied Zoology*, 84(1), 16. <https://doi.org/10.1186/s41936-023-00338-6>
71. Maria, M., Cai, R., Castillo, P., & Zheng, J. (2018). Morphological and molecular characterisation of *Hemicriconemoides paracamelliae* sp. n. (Nematoda: Criconematidae) and two known species of *Hemicriconemoides* from China. *Nematology*, 20(5), 403-422. <https://doi.org/10.1163/15685411-00003147>
72. Khan, M. R., Phani, V., Chauhan, K., Somvanshi, V. S., Pervez, R., & Walia, R. K. (2019). Redescription and molecular characterisation of *Hemicriconemoides rosae* Rathour, Sharma, Singh & Ganguly, 2003 from rhizosphere of sugarcane in India. *Nematology*, 21(7), 767-778. <https://doi.org/10.1163/15685411-00003251>
73. Li, J., Zou, C., Xu, J., Ji, X., Niu, X., Yang, J., ... & Zhang, K. Q. (2015). Molecular mechanisms of nematode-nematophagous microbe interactions: basis for biological control of plant-parasitic nematodes. *Annual review of phytopathology*, 53, 67-95. <https://doi.org/10.1146/annurev-phyto-080614-120336>
74. Khanam, S. (2016). Characterisation of the interaction between rice and the parasitic nematode *Ditylenchus angustus* (Doctoral dissertation, Ghent University). <https://doi.org/10.1163/15685411-00003251>

75. Geraert, E. (2010). The Criconematidae of the world: identification of the family Criconematidae (Nematoda). Academia press. [https://books.google.co.in/books?hl=en&lr=&id=mnfidGUVi-toC&oi=fnd&pg=PA4&dq=63.%09Geraert,+E.+\(2010\).+The+Criconematidae+of+the+world:+identifica-tion+of+the+family+Criconematidae+\(Nematoda\).+Academia+press&ots=5-oFCgh-CHy&sig=eOM0ovyrgFoabtyaVaZpoS6lgGY&redir\\_esc=y#v=onepage&q=63.%09Gera-ert%2C%20E.%20\(2010\).%20The%20Criconematidae%20of%20the%20world%3A%20identifica-tion%20of%20the%20family%20Criconematidae%20\(Nematoda\).%20Academia%20press&f=false](https://books.google.co.in/books?hl=en&lr=&id=mnfidGUVi-toC&oi=fnd&pg=PA4&dq=63.%09Geraert,+E.+(2010).+The+Criconematidae+of+the+world:+identifica-tion+of+the+family+Criconematidae+(Nematoda).+Academia+press&ots=5-oFCgh-CHy&sig=eOM0ovyrgFoabtyaVaZpoS6lgGY&redir_esc=y#v=onepage&q=63.%09Gera-ert%2C%20E.%20(2010).%20The%20Criconematidae%20of%20the%20world%3A%20identifica-tion%20of%20the%20family%20Criconematidae%20(Nematoda).%20Academia%20press&f=false)
76. Mirghasemi, N., Fanelli, E., Vovlas, A., Troccoli, A., Jamali, S., & De Luca, F. (2024). First Report of Hemicriconemoides kanayaensis (Nematoda: Criconematidae) on Tea Plantations in Iran. *Journal of Nematology*, 56(1), 20240044. <https://doi.org/10.2478/jofnem-2024-0044>
77. Takagi, K. (1969) Nematodes confronts tea plantations in Japan. *Japanese Agricultural Research Quarterly* 4, 27–32. <https://www.jircas.go.jp/en/publication/jarq/4/1-27-32>
78. McSorley, R., Campbell, C. W., & Goldweber, S. (1981). Observations on a mango decline in south Florida. In *Proceedings of the... annual meeting of the Florida State Horticultural Society* (Vol. 93). <https://ufdc.ufl.edu/UF00097625/00093/284>

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## НЕМАТОДИ У ВИРОЩУВАННІ ЧАЮ: ПАТОГЕНЕЗ ТА ЗАХИСНІ РЕАКЦІЇ РОСЛИН (ОГЛЯД)

**Мета.** Визначити складний зв'язок між чайними рослинами та важливими рослинними паразитичними нематодами, зосереджуючись на серйозній шкоді, яку ці шкідники завдають корінню, також складні захисні стратегії, що використовуються чайними рослинами, зокрема їхній синтез захисних вторинних метаболітів для боротьби з атаками нематод.

**Результати.** Фітопаразитичні нематоди становлять серйозну економічну загрозу для *Camellia sinensis*, найважливішої чайної культури у світі, що призводить до втрат врожаю на 11–55% та до 1 мільярда доларів щорічно. Затримка росту, в'янення та зниження врожаю чаю спричинені близько 80 видами нематод, які пошкоджують коріння, включаючи *Pratylenchus*, *Radopholus*, *Meloidogyne* та *Hemicriconemoides*. Через свою нерухомість чайні рослини розвинули складні захисні механізми. До них належать утворення нематоцидних вторинних метаболітів (поліфенолів, алкалоїдів, терпеноїдів), системні сигнальні шляхи, що запускаються фітогормонами (етилен, жасмонат та саліцилова кислота), що активують захисні гени, та фізичні бар'єри, такі як лігнін та суберин у клітинних стінках. Контроль нематод також здійснюється корисними ґрунтовими бактеріями.

**Висновки.** Для сталого управління важливо розуміти взаємозв'язки. Щоб забезпечити економічне майбутнє чайного бізнесу, майбутні дослідження повинні використовувати природні захисні механізми для покращення комплексної боротьби зі шкідниками.

**КЛЮЧОВІ СЛОВА:** *Camellia sinensis*, рослинні паразитичні нематоди, захист рослин, вторинні метаболіти, інтегрована боротьба зі шкідниками

#### ***Конфлікт інтересів***

Автори заявляють, що конфлікту інтересів щодо публікації цього рукопису немає. Крім того, автори повністю дотримувались етичних норм, включаючи плагіат, фальсифікацію даних та подвійну публікацію.

**Внесок авторів:** всі автори зробили рівний внесок у цю роботу

У цьому дослідженні штучний інтелект використовувався тільки для пошуку та компіляції актуальної наукової літератури за цією темою. Автори, несуть повну відповідальність за інтелектуальний зміст роботи, особисто виконували найважливіші завдання аналізу даних, синтезу інформації та формулювання наукових висновків.

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