БІОФІЗИЧНИЙ ВІСНИК, 2023, Вип. 50

MOLECULAR BIOPHYSICS

Original article

https://doi.org/10.26565/2075-3810-2023-50-04

UDC 577.3.01; 577.38; 577.3.001.57

NEW PROMISING AGENTS AGAINST COPD AND ASTHMA AMONG THE AMIDES OF 1-OXO-3-PHENYL-ISOCHROMAN-6-CARBOXYLIC ACID

Alex Nyporko¹, Olga Tsymbalyuk¹, Ivan Voiteshenko¹, Sergiy Starosyla², Mykola Protopopov³, Volodymyr Bdzhola⁴

¹ Taras Shevchenko National University of Kyiv, 64/13 Volodymyrska St., Kyiv, 01601, Ukraine;

² Receptor.AI Inc., 20–22 Wenlock Road, London, N1 7GU, United Kingdom;

³ Chemspace LLC, 85, office 1 Winston Churchill St., Kyiv, 02094, Ukraine;

⁴ Institute of Molecular Biology and Genetics of NAS of Ukraine, 150 Zabolotnogo St., Kyiv, 03143, Ukraine e-mail: <u>a nyporko@knu.ua</u>

Submitted December 3, 2023; Revised December 12, 2023;

December 14, 2023

Background: Bronchodilators, which are compounds that can relax airway smooth muscle, are perhaps the most important component of combination therapy for chronic obstructive pulmonary disease, one of the most common non-communicable diseases in the world, which is the second most lethal disease after cardiovascular disease. Unfortunately, current clinical bronchodilators, whose activity is mediated by their interaction with muscarinic acetylcholine receptors, have side effects (up to myocardial infarction) due to their cross-affinity for different types of these receptors, including those prevalent in the heart muscle.

Objectives: The aim of this work is to search/develop compounds — effective bronchodilators capable of selectively inhibiting type 3 muscarinic acetylcholine receptors (M_3 receptors), predominantly present in smooth muscles and not characteristic of cardiomyocytes.

Materials and Methods: High-throughput virtual screening of a collection of 150,000 compounds was conducted on the spatial structure of the M_3 receptor, reconstructed in our previous studies. The effect of substances on contractile activity was investigated using tensometry in isometric mode on multicellular tracheal preparations. Antagonistic activity and type of inhibition were determined against the background of acetylcholine application (concentration range 10^{-10} – 10^{-3} M). To establish the affinity value of the compound-antagonist, the Schild regression equation was used.

Results: Based on virtual screening data, a series of compounds — amides of 1-oxo-3-phenyl-iso-chroman-6-carboxylic acid — were selected for biological testing. For two of these compounds (Compounds 1 and 7), the ability to selectively inhibit M_3 receptors was demonstrated. Specifically, the affinity value pK_B for Compound 1 was 7.28 ± 0.70 , with an IC₅₀ of $5.25 \cdot 10^{-8}$ M. A critically important advantage of this compound is its ability, at equal concentrations, to more effectively inhibit signal transmission through M_3 receptors compared to ipratropium bromide — a clinical cholinergic receptor inhibitor.

Conclusions: The sufficient effectiveness of inhibition and significantly increased selectivity of the studied compounds specifically towards M_3 receptors provide strong grounds to consider these compounds as promising precursors of new generation cholinolytic drugs with targeted action on M_3 -type cholinergic receptors.

KEY WORDS: chronic obstructive pulmonary disease (COPD); acetylcholine receptor; virtual screening; molecular docking; tensometry; selective M3 antagonists.

In cites: Nyporko A, Tsymbalyuk O, Voiteshenko I, Starosyla S, Protopopov M, Bdzhola V. New promising agents against COPD and asthma among the amides of 1-oxo-3-phenyl-isochroman-6-carboxylic acid. Biophysical Bulletin. 2023;50:36–47. https://doi.org/10.26565/2075-3810-2023-50-04

Open Access. This article is licensed under a Creative Commons Attribution 3.0 <u>http://creativecommons.org/licenses/by/3.0/</u>

© Nyporko A., Tsymbalyuk O., Voiteshenko I., Starosyla S., Protopopov M., Bdzhola V., 2023.

In the human respiratory tract, M_2 and M_3 type cholinergic receptors play a direct role in regulating the lumen of the bronchial tree [1–4]. The physiological functions of M_2 cholinergic receptors, located in smooth muscle cells and postganglionic parasympathetic neurons, are to limit β_2 -adrenoreceptor-mediated relaxation of smooth muscles by reducing intracellular cAMP concentration and to suppress the release of the neurotransmitter acetylcholine from nerve endings of postganglionic neurons [1, 5, 6]. Although M_2 type receptors significantly outnumber M_3 cholinergic receptors in smooth muscle cell membranes, the latter are functionally the main type of acetylcholine receptors that facilitate their contraction and narrowing of the respiratory tract lumen, as well as regulate the secretory activity of mucosal glands [1, 2, 7–9].

Asthma and Chronic Obstructive Pulmonary Disease (COPD) are among the most common non-infectious diseases worldwide. According to data from the Global Asthma Network and the Global Initiative for Asthma, asthma affects 1 to 18% of the population in various countries, with a total global number of patients exceeding 300 million; a concerning global trend is the annual increase in patients diagnosed with asthma by 2.9% [9]. Asthma is characterized by transient airflow limitation in the airways, as well as airway hyperresponsiveness and sometimes remodeling of the airway wall tissues; these symptoms are also primary in COPD.

Pharmacological drugs used in the treatment of asthma and COPD are divided into several groups: long-acting β_2 -adrenoceptor agonists (LABA), short-acting β_2 -adrenoceptor agonists (SABA), long-acting muscarinic acetylcholine receptor antagonists (LAMA), short-acting muscarinic acetylcholine receptor antagonists (SAMA), as well as inhaled and oral corticosteroids.

Currently, the most commonly used drug in Ukraine for COPD and bronchial asthma with M-cholinolytic activity is ipratropium bromide [10, 11]. Like atropine, ipratropium is a competitive antagonist of muscarinic acetylcholine receptors. It has gained widespread use in the therapy of asthma and COPD, as unlike atropine, it does not cross the blood-brain barrier and is poorly absorbed through the gastrointestinal tract walls. Ipratropium is recommended by the Global Initiative for Chronic Obstructive Lung Disease 2009 (according to the Global strategy for diagnosis, management, and prevention of COPD) for four times daily inhalation use by COPD patients as a short-acting muscarinic antagonist (SAMA), with the maximum effect achieved within 60–90 minutes and the duration of therapeutic effect lasting 4–6 hours.

However, the optimal strategy for the therapy of obstructive phenomena in the airways with cholinolytic drugs (more precisely, drugs with properties of long-acting muscarinic antagonist — LAMA) involves the use of M₃-selective antagonists [1, 12]. Current COPD therapy protocols (USA, EU countries) and asthma include the use of such LAMA: tiotropium (recommended at a dose of 5 or 18 μ g depending on the nebulizer functioning method, once daily) and aclidinium (recommended at a dose of 400 μ g twice daily), glycopyrronium (recommended at a dose of 50 μ g once daily), and umeclidinium (for COPD treatment and potential use in asthma treatment, recommended at a dose of 62.5 μ g once daily) [2, 13]. According to a meta-analysis [14], for newer LAMA drugs (aclidinium, glycopyrronium, and glycopyrronium), no significant therapeutic advantages were found compared to the earlier tiotropium bromide (the only LAMA drug for COPD therapy until 2012).

Thus, currently, the active substance in drugs for the therapy of uncontrolled asthma is tiotropium bromide — a structural analog of ipratropium bromide, which is a functional antagonist of M_3 cholinergic receptors. This is because, although it binds to M_2 receptors, it dissociates from these significantly faster than from M_3 type receptors. Compared to ipratropium, tiotropium bromide has lower oral bioavailability and systemic side effects, and an extended half-life in patient's body [15–17]. However, the use of tiotropium bromide for the

therapy of stable asthma may be ineffective due to insufficient drug dosage, while increasing the dosage can lead to side effects [1, 18].

Therefore, the aim of our study was the directed search and biological testing of lowmolecular-weight selective inhibitors of M₃ type muscarinic acetylcholine receptors.

MATERIALS AND METHODS

Virtual Screening

The predicted structure of the M₃-type muscarinic acetylcholine receptor [19] was used as a target for virtual screening. Two binding sites were identified. Binding site 1 (orthosteric) and binding site 2 (allosteric) utilizing the ICM Pocket Finder tool [20]. The first site is a combined pocket from the area located in a fold formed by helically twisted transmembrane domains, with which acetylcholine binds, as well as the area located higher, between the loops that bind the transmembrane domains. The second is on the reverse, intracellular part, formed by transmembrane domains. The orthosteric site has better characteristics than the allosteric one (larger volume of 1013.7 Å³ compared to 461.8 Å³ for the allosteric site, aromatic factor of 0.15 compared to 0, and drug-like index (DLID) [21] of 1.35 compared to 0.13). After detailed analysis and determination of the parameters of these sites, the orthosteric site was chosen for docking studies.

Virtual screening of the Department of Medicinal Chemistry Institute of Molecular Biology and Genetics NAS of Ukraine compounds collection (150,000 compounds) was conducted utilizing AutoDock 4.2 [22] molecular docking software with previously reported parameters and protocols [23]. 3,000 top-scored compounds (2% of all screened compounds) were selected according to AutoDock score for visual inspection and cherry-picking.

The BIOVIA Discovery Studio Visualizer [24] was utilized for visual inspection and ligand-target interaction analysis of top scored compounds. The parameters of the visualizer related to the construction of molecular bonds were used by default. The criteria for selecting compounds for pharmacological tests were as follows: 1. Compounds should bind to the acetylcholine-binding site; 2. Compounds should also bind to the subpocket above the acetylcholine-binding site for enhanced affinity and selectivity; 3. There must be at least one hydrogen bond.

Functional pharmacological studies of compound activity in vitro

Testing the effect of muscarinic acetylcholine receptor inhibitor substances was carried out on the contractile activity of the smooth muscles of the trachea using male Wistar rats (weighing within 230–250 g). Rats were euthanized under ether anesthesia, and all animal manipulations were conducted in accordance with the International Convention on the Work with Animals and the Law of Ukraine 'On the Protection of Animals from Cruel Treatment' (protocol of the meeting of the bioethics commission of the National Scientific Center 'Institute of Biology and Medicine' No. 3 dated May 2, 2019).

The study of contractile activity was conducted using tensometry in isometric mode on multicellular tracheal preparations, which were rings with an intact mucosal lining containing at least four cartilaginous rings. For the experiments, tracheal fragments from the beginning of the bifurcation to 1-1.5 cm above the bifurcation were used.

In the experiments, Krebs solution was used (mM): 120.4 NaCl; 5.9 KCl; 15.5 NaHCO₃; 1.2 NaH₂PO₄; 1.2 MgCl₂; 2.5 CaCl₂; 11.5 glucose; the pH of the solution was 7.4. Tracheal rings were placed in a working chamber (volume 2 ml) bubbled with an oxygenated gas mixture of Krebs solution (95% O₂/5% CO₂, flow rate — 5 ml/min), and thermostated at 37°C. The tracheal preparations were given a passive tension of 10 mN and left for 1 hour; the study began after recording several reproducible contractile reactions to the application of hyperkalemic solution and acetylcholine (10^{-6} M). The contractile activity was investigated using a capacitive force sensor; the amplified signal was recorded using an ADC.

Preliminary screening of compounds for their ability to inhibit muscarinic cholinergic receptors was conducted by recording contractile responses of tracheal preparations to the application of acetylcholine (10^{-5} M) under the condition of prior action of the tested compounds (at a concentration of 10^{-4} M) for 10 minutes.

To study the antagonistic activity of muscarinic cholinergic receptor antagonists (ipratropium bromide and tested substances), contractile responses of tracheal preparations to the application of acetylcholine at a concentration of 10^{-4} M were recorded under cumulative increases in antagonist concentrations (10^{-10} – 10^{-5} M, with the antagonist's prior action time of 10 minutes) and concentration-effect curves were constructed, from which the IC₅₀ value was determined; the maximum contractile response (100%) was considered as the contraction to the application of acetylcholine without an antagonist.

To confirm the competitive type of inhibition by ipratropium bromide and tested substances, as well as to study their functional antagonism, concentration-effect curves of acetylcholine were recorded (acetylcholine concentration range 10^{-10} – 10^{-3} M). Subsequently, the Schild regression equation was used to determine the affinity of the compound-antagonist. According to the Schild method, the antagonist-induced parallel shift of the concentration-effect curves was determined as the ratio of equieffective concentrations (CR) of the agonist in control and in the presence of an antagonist. From the Schild plots, the functional antagonism index (affinity index of the competitive antagonist) pK_B and the tangent of the slope angle were determined [25–27].

To study the cellular mechanisms of action of the compounds, contractile responses of the trachea to the application of the selective mAChR₃ agonist cevimeline (10^{-4} M), effectors of adrenoreceptors (adrenaline, 10^{-5} M; propranolol, 10^{-5} M; isoproterenol, 10^{-5} M), and nicotinic cholinergic receptors (nicotine, 10^{-4} M), blocker of inositol 1,4,5-trisphosphate-sensitive (IP₃) Ca²⁺ channels of the sarcoplasmic reticulum (2-APB, 10^{-4} M), and phospholipase C inhibitor (U-73122, 10^{-6} M) were recorded.

All compounds under study were initially dissolved in DMSO and introduced into the smooth muscle preparations in such a way that the final concentration of DMSO was 0.1%. Additionally, all experiments were conducted in the presence of DMSO in the bathing solution at a concentration of 0.1%.

Experimental data were analyzed using Origin Pro 2018 software. The samples were tested for their adherence to normally distributed populations using the Shapiro-Wilk criterion. To determine significant differences between the mean values of the samples, a paired t-test was used; multiple comparisons were conducted using one-way analysis of variance (ANOVA). In all cases, results were considered significant if p < 0.05. The significance of data approximation by a linear function was analyzed using Fisher's F-criterion; the coefficients of determination (R^2) were not less than 0.9. Results are presented as mean \pm standard error of mean (SEM), n – number of experiments.

RESULTS AND DISCUSSION

For pharmacological testing *in vitro* on multicellular smooth muscle preparations of rat tracheal rings, seven most promising compounds — amides of 1-oxo-3-phenyl-isochroman-6-carboxylic acid (1–7, Table 1) with predicted cholinolytic activity were selected through preliminary *in silico* screening. As a target for screening, the spatial structure of the M_3 type cholinergic receptor, reconstructed in our previous studies [19], were used. Structure of these compounds are presented in Table 1.



 Table 1. Structures and physico-chemical properties of the studied amides of 1-oxo-3-phenyl-isochroman-6-carboxylic acid

In the first series of experiments, the reaction of smooth muscle preparations (SMP) of the trachea to the application of acetylcholine (10^{-5} M) was tested under the condition of prior

action of the tested compounds (all at a concentration of 10^{-4} M) for 10 minutes. Thus, two compounds were identified with the ability to suppress acetylcholine-induced contractions, identified as 1 and 7 (Fig. 1).



Fig. 1. Changes in the amplitude (F_{max}) of acetylcholine-induced (10^{-5} M) contractions of rat tracheal preparations by compounds (used at a concentration of 10^{-4} M, pre-incubation time 10 min).

Control values were taken as 100% (n = 7).

** — p < 0.01 and * — p < 0.05 — significant difference compared to control.

Spatial structures of the corresponding complexes of active compounds are depicted in Fig. 2. Both compounds interact with the M_3 cholinergic receptor in the same manner. The 1-oxo-3-phenyl-isochroman moiety interacts within the acetylcholine binding site, specifically forming a series of hydrophobic interactions with Tyr148, Trp503, Tyr506, Tyr529, Cys532, and a hydrogen bond with Asn507. The NH of amide group forms two hydrogen bonds with Tyr148 and Tyr506. The o-tolyl group of compound **7** engages in hydrophobic interactions with Ile222 and Leu225. Methyl o-benzoate group of compounds **1** has additional hydrophobic interactions with Leu144 and Tyr148, as well as hydrogen bonds with Tyr148, Ile222, and Tyr529.

In the subsequent series of experiments, acetylcholine-induced contractions of rat tracheal preparations were recorded under the action of selected compounds 1 and 7 (all used at a concentration of 10^{-4} M) against the background of prior incubation of SMP with the known cholinergic receptor antagonist ipratropium (used at a concentration of 10^{-6} M). Since the tested compounds did not affect the baseline tension and contractile response of SMP in the presence of ipratropium, it can be asserted that these substances are tropic to muscarinic cholinergic receptors.

The mentioned compounds did not affect the SMP responses to the application of the nicotinic cholinergic receptor agonist nicotine (used concentration 10^{-6} M), supporting the hypothesis of the selective action of the tested substances specifically on muscarinic acetylcholine receptors [21].



Fig. 2. Molecular complexes of M_3 cholinergic receptor with compound 1 (A) and compound 7 (B) obtained by molecular docking. Hydrogen bonds are shown by the green dotted lines, hydrophobic interactions are indicated by magenta dotted lines.

It was also established that both compounds and the known non-selective inhibitor of muscarinic cholinergic receptors, ipratropium bromide (in all cases used at a concentration of 10^{-6} M, with a pre-incubation duration of 10 minutes in the presence of the tested substance), could inhibit the contractions of SMP of the trachea activated by the selective M₃ cholinergic

receptor agonist cevimeline (fixed concentration of 10^{-4} M). The greatest inhibitory properties against M₃ cholinergic receptors were observed for compound **7** — under these conditions, cevimeline-induced contraction was $24.5 \pm 3.6\%$ relative to the control, taken as 100% (n = 7, p < 0.001). For comparison, a similar effect on cevimeline-induced SMP tracheal contractions for ipratropium bromide was registered, a compound that is the active ingredient in medicinal products for the therapy of bronchial asthma and COPD. It was found that ipratropium bromide at a concentration of 10^{-6} M causes an average inhibition of cevimeline-induced contraction to $63.2 \pm 5.1\%$ relative to the control, taken as 100% (n = 7, p < 0.01).

Subsequently, for the compound 7, 'acetylcholine concentration-effect' curves were registered and analyzed, the type of inhibition, affinity indicators, and IC₅₀ were determined. It was found that the slope of the Schild regression line (0.88 ± 0.12 , coefficient of determination R² = 0.98) (Fig. 3, 4) indicates a competitive mechanism of action of this substance. The affinity value of this compound pK_B was 7.28 ± 0.70 and IC₅₀ = $5.25 \cdot 10^{-8}$ M.



Fig. 3. 'Acetylcholine concentration-effect' curves in the presence of compound **7** for the activation of smooth muscle contractions of rat trachea: curve 1 — control (concentration range 10^{-10} – 10^{-3} M), curves 2–4 — against the background of **7** at concentrations of $5 \cdot 10^{-7}$ M, 10^{-6} M, and 10^{-5} M, respectively. The amplitude of contractions under the action of antagonists is recalculated in % compared to acetylcholine-induced contraction in control (10^{-4} M), taken as 100%. Data are presented as M ± SEM, n = 7.

It has also been established that compound **7** does not significantly affect the nicotinic cholinergic receptors and adrenergic receptors of the respiratory tract in rats. Using a phospholipase C inhibitor (U-73122) and blockers of inositol 1,4,5-trisphosphate-sensitive (2-APB) and ryanodine-sensitive (caffeine) Ca^{2+} channels of the sarcoplasmic reticulum, it was demonstrated that the aforementioned compounds act on the intracellular signaling cascade through M₃ type muscarinic cholinergic receptors [28–31].



Fig. 4. Schild plot for the action of compound 7. Data are presented as $M \pm SEM$, n = 7.

Previously, we established [19] that for the known non-selective competitive antagonist of muscarinic cholinergic receptors, ipratropium bromide, the slope of the Schild regression line was 0.79 ± 0.07 (coefficient of determination $R^2 = 0.99$), and the affinity value pK_B was 9.14 ± 0.62 with an IC₅₀ of $7.24 \cdot 10^{-10}$ M. Therefore, compound **7** has a lower affinity and is characterized by higher EC₅₀ values compared to ipratropium bromide. However, a critically important advantage of compound **7** is its ability, at equal concentrations, to more effectively inhibit signal transmission through M₃ cholinergic receptors compared to ipratropium bromide.

CONCLUSIONS

Biological testing was conducted on smooth muscle preparations (SMP) of the trachea for compounds in the group of amides of 1-oxo-3-phenyl-isochroman-6-carboxylic acid with predicted inhibitory activity towards mAChRs: their pharmacological effects and parameters were studied. It was found that the compound **7** effectively inhibits (with an average IC₅₀ value of $5.25 \cdot 10^{-8}$ M) and at equal concentrations significantly more inhibits signal transduction specifically through M₃ cholinergic receptors compared to ipratropium bromide, without having a significant effect on M₂ cholinergic receptors. Therefore, there are all reasons to consider these compounds as promising precursors of new generation cholinolytic drugs with targeted action on M₃ cholinergic receptors.

ACKNOWLEDGEMENTS

The study was funded by the National Research Foundation of Ukraine, Project 2020.01/0543.

CONFLICT OF INTEREST

The authors report that there is no conflict of interest.

Authors' ORCID ID

Alex Nyporko D https://orcid.org/0000-0003-1664-6837 Olga Tsymbalyuk D https://orcid.org/0000-0002-4524-7627 Ivan Voiteshenko D <u>https://orcid.org/0000-0003-2434-9218</u> Sergiy Starosyla D <u>https://orcid.org/0000-0002-5103-0635</u> Mykola Protopopov D <u>https://orcid.org/0000-0002-2716-4844</u> Volodymyr Bdzhola D https://orcid.org/0000-0003-0315-450X

REFERENCES

- Cavazos Galván M. Asthma in emergency department. Guidelines, physicians and patients. Rev Alerg Mex. 2006 Jul-Aug;53(4):136–43. Available from: <u>https://pubmed.ncbi.nlm.nih.gov/17137189/</u>
- Zeng Z, Mukherjee A, Varghese AP, Yang XL, Chen S, Zhang H. Roles of G protein-coupled receptors in inflammatory bowel disease. World J Gastroenterol. 2020 Mar 28;26(12):1242–61. https://doi.org/10.3748/wjg.v26.i12.1242
- Ilyaskina OS, Lemoine H, Bünemann M. Lifetime of muscarinic receptor–G-protein complexes determine coupling efficiency and G-protein subtype selectivity. Proc Natl Acad Sci. 2018 May 8;115(19):5016–21. <u>https://doi.org/10.1073/pnas.1715751115</u>
- Saternos HC, Almarghalani DA, Gibson HM, Meqdad MA, Antypas RB, Lingireddy A, et al. Distribution and function of the muscarinic receptor subtypes in the cardiovascular system. Physiol Genomics. 2018 Jan 1;50(1):1–9. <u>https://doi.org/10.1152/physiolgenomics.00062.2017</u>
- 5. Ishii M, Kurachi Y. Muscarinic Acetylcholine Receptors. Curr Pharm Des. 2006 Oct 1;12(28):3573-81. https://doi.org/10.2174/138161206778522056
- Tobin G, Giglio D, Lundgren O. Muscarinic receptor subtypes in the alimentary tract. J Physiol Pharmacol. 2009 Mar;60(1):3–21. Available from: <u>https://europepmc.org/article/med/19439804</u>
- Lee HW, Park J, Jang EJ, Lee CH. Comparisons of exacerbations and mortality among LAMA/LABA combinations in stable chronic obstructive pulmonary disease: systematic review and Bayesian network meta-analysis. Respir Res. 2020 Dec 25;21(1):310. <u>https://doi.org/10.1186/s12931-020-01540-8</u>
- Kruse AC, Kobilka BK, Gautam D, Sexton PM, Christopoulos A, Wess J. Muscarinic acetylcholine receptors: novel opportunities for drug development. Nat Rev Drug Discov. 2014 Jul 6;13(7):549–60. <u>https://doi.org/10.1038/nrd4295</u>
- Rhee CK, Yoshisue H, Lad R. Fixed-Dose Combinations of Long-Acting Bronchodilators for the Management of COPD: Global and Asian Perspectives. Adv Ther. 2019 Mar 11;36(3):495–519. https://doi.org/10.1007/s12325-019-0893-3
- Abrams P, Andersson K, Buccafusco JJ, Chapple C, de Groat WC, Fryer AD, et al. Muscarinic receptors: their distribution and function in body systems, and the implications for treating overactive bladder. Br J Pharmacol. 2006 Jul 29;148(5):565–78. <u>https://doi.org/10.1038/sj.bjp.0706780</u>
- Eglen RM. Muscarinic Receptor Subtype Pharmacology and Physiology. In: King FD, Lawton G, editors. Progress in Medicinal Chemistry. Vol. 43. Elsevier; 2005. p. 105–36. <u>https://doi.org/10.1016/s0079-6468(05)43004-0</u>
- 12. Barnes PJ. Muscarinic receptor subtypes in airways. Life Sci. 1993 Jan;52(5-6):521-7. https://doi.org/10.1016/0024-3205(93)90310-y
- 13. Miravitlles M, Anzueto A, Jardim JR. Optimizing bronchodilation in the prevention of COPD exacerbations. Respir Res. 2017 Dec 20;18(1):125. <u>https://doi.org/10.1186/s12931-017-0601-2</u>
- 14. Eglen RM. Overview of Muscarinic Receptor Subtypes. In: Muscarinic Receptors. Berlin, Heidelberg: Springer Berlin Heidelberg; 2012. p. 3–28. <u>https://doi.org/10.1007/978-3-642-23274-9_1</u>
- 15. Gomes F, Cheng SL. Pathophysiology, Therapeutic Targets, and Future Therapeutic Alternatives in COPD: Focus on the Importance of the Cholinergic System. Biomolecules. 2023 Mar 5;13(3):476. https://doi.org/10.3390/biom13030476
- Maqsood MH, Rubab K, Maqsood MA. The Role of Revefenacin in Chronic Obstructive Pulmonary Disease. Cureus. 2019 Apr 10; 11(4):e4428. <u>https://doi.org/10.7759/cureus.4428</u>
- 17. Nadler LS, Rosoff ML, Hamilton SE, Kalaydjian AE, McKinnon LA, Nathanson NM. Molecular analysis of the regulation of muscarinic receptor expression and function. Life Sci. 1999 Jan;64(6–7):375–9. https://doi.org/10.1016/s0024-3205(98)00577-3
- Zaagsma J, Roffel AF, Meurs H. Muscarinic control of airway function. Life Sci. 1997 Feb;60(13–14):1061– 8. <u>https://doi.org/10.1016/s0024-3205(97)00048-9</u>
- Nyporko A, Tsymbalyuk O, Voiteshenko I, Starosyla S, Protopopov M, Bdzhola V. Computer-aided design of muscarinic acetylcholine receptor M3 inhibitors: Promising compounds among trifluoromethyl containing hexahydropyrimidinones/thiones. Mol Inform. 2023 Aug 9;42(8–9). <u>https://doi.org/10.1002/minf.202300006</u>
- Abagyan R, Raush E, Totrov M. ICM Manual v.3.9 [Internet]. Molsoft, LLC. [cited 2023 Nov 30]. Available from: <u>https://www.molsoft.com/icm</u>

- 21. Protopopov M V., Starosyla SA, Borovykov O V., Sapelkin VN, Bilokin Y V., Bdzhola VG, et al. Hit identification of CK2 inhibitors by virtual screening. Biopolym Cell. 2017 Aug 31;33(4):291–301. https://doi.org/10.7124/bc.00095B
- 22. Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS, et al. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. J Comput Chem. 2009 Dec 27;30(16):2785–91. <u>https://doi.org/10.1002/jcc.21256</u>
- 23. Sheridan RP, Maiorov VN, Holloway MK, Cornell WD, Gao YD. Drug-like Density: A Method of Quantifying the "Bindability" of a Protein Target Based on a Very Large Set of Pockets and Drug-like Ligands from the Protein Data Bank. J Chem Inf Model. 2010 Nov 22;50(11):2029–40. <u>https://doi.org/10.1021/ci100312t</u>
- 24. Dassault Systèmes. BIOVIA Discovery Studio Visualizer [Internet]. Dassault Systèmes. 2020 [cited 2023 Nov 30]. Available from: <u>https://discover.3ds.com/discovery-studio-visualizer-download</u>
- 25. Tsymbaliuk OV, Naumenko AM, Skoryk MA, Nyporko OYu, Davidovska TL, Skryshevsky VA. Histamineand nicotine-stimulated modulations of mechanic activity of smooth muscles in gastrointestinal tract at the impact of nanosized TiO₂ material. Biopolym Cell. 2016 Apr 30;32(2):140–9. <u>https://doi.org/10.7124/bc.000917</u>
- 26. Naumenko AM, Dmytrenko O V., Shapoval LM, Tsymbalyuk O V., Sagach VF, Davydovska TL. Effects of Injections of Nanostructured Titanium Dioxide into the Rat Medullary Nuclei Involved in Cardiovascular Control. Neurophysiology. 2018 Nov 18;50(6):409–14. <u>https://doi.org/10.1007/s11062-019-09772-1</u>
- 27. Naumenko AM, Shapoval LM, Nyporko AYu, Voiteshenko MI, Tsymbalyuk A V., Sagach VF, et al. Computer Simulation of Molecular Interaction Between Baclofen and the GABA_B Receptor. Neurophysiology. 2017 Feb 1;49(1):2–7. <u>https://doi.org/10.1007/s11062-017-9623-0</u>
- 28. Luo L, Zhang G, Mao L, Wang P, Xi C, Shi G, et al. Group II muscarinic acetylcholine receptors attenuate hepatic injury via Nrf2/ARE pathway. Toxicol Appl Pharmacol. 2020 May;395:114978. https://doi.org/10.1016/j.taap.2020.114978
- 29. Baroffio M, Crimi E, Brichetto L, Zappi L, Rehder K, Brusasco V. Pre-junctional muscarinic autoreceptors in bovine airways. Respir Physiol Neurobiol. 2012 Jan;180(1):45–51. <u>https://doi.org/10.1016/j.resp.2011.10.007</u>
- 30. Naumenko AM, Nyporko AYu, Tsymbalyuk OV, Golius A, Shapoval LM, Davidovska TL. 3D reconstruction of a full-size GABA_B receptor. Neurophysiology. 2015;47:364–75. <u>https://doi.org/10.1007/s11062-016-9544-</u> <u>3</u>
- 31. Aparici M, Carcasona C, Ramos I, Montero JL, Otal R, Ortiz JL, et al. Pharmacological Profile of AZD8871 (LAS191351), a Novel Inhaled Dual M₃ Receptor Antagonist/β₂-Adrenoceptor Agonist Molecule with Long-Lasting Effects and Favorable Safety Profile. J Pharmacol Exp Ther. 2019 Jul;370(1):127–36. https://doi.org/10.1124/jpet.118.255620

НОВІ ПЕРСПЕКТИВНІ ЗАСОБИ ПРОТИ ХОЗЛ ТА АСТМИ СЕРЕД АМІДІВ 1-ОКСО-3-ФЕНІЛ-ІЗОХРОМАН-6-КАРБОНОВОЇ КИСЛОТИ

Олексій Нипорко¹, Ольга Цимбалюк¹, Іван Войтешенко¹, Сергій Старосила², Микола Протопопов³, Володимир Бджола⁴

¹ Київський національний університет імені Тараса Шевченка, вул. Володимирська, 64/13, Київ, 01601, Україна; ² Receptor.AI Inc., буд. 20, поверх 22, вул. Венлок, Лондон, N1 7GU, Об'єднане Королівство;

³ Chemspace LLC, вул. Вінстона Черчилля, 85, офіс 1, Київ, 02094, Україна;

⁴ Інститут молекулярної біології і генетики НАН України, вул. Заболотного, 150, Київ, 03143, Україна e-mail: <u>a nyporko@knu.ua</u>

Надійшла до редакції 3 грудня 2023 р. Переглянута 12 грудня 2023 р.

Прийнята до друку 14 грудня 2023 р.

Актуальність. Бронходилататори — сполуки, здатні розслабляти гладеньку мускулатуру повітроносних шляхів, є чи найважливішим компонентом комбінованої терапії хронічного обструктивного захворювання легень — одного з найбільш поширених у світі неінфекційних захворювань, що займає друге місце за летальністю після серцево-судинних захворювань. На жаль, сучасні клінічні бронходилататори, чия активність опосередкована їх взаємодією з мускариновими рецепторами ацетилхоліну, мають побічні ефекти (до інфаркту міокарда) внаслідок їх перехресної спорідненості до різних типів цих рецепторів, зокрема, і до тих, що розповсюджені в серцевому м'язі.

Мета роботи: пошук/розробка сполук — ефективних бронходилататорів, здатних селективно інгібувати мускаринові рецептори ацетилхоліну типу 3 (М₃-рецептори), які представлені переважно в гладеньких м'язах і не характерні для кардіоміоцитів.

Матеріали і методи. Високопродуктивний віртуальний скринінг колекції 150000 сполук було здійснено щодо просторової структури М3-рецептора, реконструйованого в наших попередніх дослідженнях. Вплив речовин на скорочувальну активність досліджували методом тензометрії у ізометричному режимі на мультиклітинних препаратах трахеї. Антагоністичну активність і тип інгібування визначали на фоні аплікування ацетилхоліну (діапазон концентрацій 10⁻¹⁰–10⁻³ М). Для встановлення величини афінності сполуки-антагоніста використовували рівняння регресії Шілда.

Результати. За даними віртуального скринінгу було обрано для біологічного тестування ряд сполук — амідів 1-оксо-3-феніл-ізохроман-6-карбонової кислоти. Для двох із них (сполуки 1 і 7) було продемонстровано здатність селективно інгібувати М₃-рецептори. Зокрема, велична афінності рК_В для сполуки 1 становила 7,28±0,70, а IC₅₀=5,25·10⁻⁸ М. Надзвичайно важливою перевагою цієї сполуки є її здатність за однакових концентрацій достовірно ефективніше пригнічувати проведення сигналу через М₃-рецептори порівняно з іпратропієм бромідом — клінічним інгібітором холінорецепторів.

Висновки. Достатня ефективність інгібування і значно підвищена селективність досліджених сполук саме стосовно М₃-рецепторів дають всі підстави вважати зазначені сполуки перспективними попередниками холінолітичних препаратів нового покоління зі спрямованою дією на холінорецептори М₃-типу.

КЛЮЧОВІ СЛОВА: хронічне обструктивне захворювання легень (ХОЗЛ); мускариновий ацетилхоліновий рецептор; віртуальний скринінг; молекулярний докінг; тензометрія; селективні МЗ-антагоністи.